NOVEL POLYETHER ANTIBIOTICS X-14873A, G AND H PRODUCED BY A *STREPTOMYCES*: TAXONOMY OF THE PRODUCING CULTURE, FERMENTATION, BIOLOGICAL AND IONOPHOROUS PROPERTIES OF THE ANTIBIOTICS

Chao-Min Liu, John W. Westley[†], T. E. Hermann, B. La T. Prosser, N. Palleroni, R. H. Evans and P. A. Miller

> Roche Research Center, Hoffmann-La Roche Inc., Nutley, NJ 07110, U.S.A.

> > (Received for publication July 17, 1986)

Novel polyether antibiotics X-14873A, X-14873G, and X-14873H are produced by the fermentation of *Streptomyces* sp. X-14873 (ATCC 31679). This report presents taxonomic studies and fermentation conditions for the antibiotic producing culture. The antibiotics are mainly active against Gram-positive bacteria. The ionophore properties of X-14873A are also characterized.

Intensive search in our laboratory^{1~4)} and others^{5~7)} in the past decade has revealed that the biosynthesis of carboxylic acid polyether antibiotics is a widespread property among streptomycetes. In this paper, we report the discovery of another novel polyether antibiotic X-14873A and its descarboxyl derivatives, X-14873G and X-14873H (Fig. 1). This report presents the taxonomy of the producing culture *Streptomyces* sp. X-14873, fermentation conditions for the antibiotic production, and biological as well as the ionophorous properties of X-14873A. The isolation and characterization of the antibiotics and other metabolites are described in the accompanying paper^{s)}.

Taxonomy of the Producing Culture

The streptomycetes culture producing antibiotic X-14873A, X-14873G, and X-14873H was isolated from a soil sample collected near a sagebrush plant in Crandall Creek, Wyoming. The culture was designated strain X-14873 and has been deposited at American Type Culture Collection where it was assigned the number ATCC 31679. *Streptomyces* sp. X-14873 also elaborates a novel actinomycin complex, of which three of the antibiotic components designated actinomycins X-14873B, X-14873C, and X-14873D have been isolated and characterized[§]. The culture has the following morphological, physiological, and chemical characteristics.

Microscopic and Macroscopic Examination

Strain X-14873 produces a substrate mycelium which does not fragment into spores, and an aerial mycelium forming *Rectus-Flexibilis* spore chains with $10 \sim 20$ spores per chain. Spores are smooth and range in size from $1.0 \times 0.52 \ \mu m$ to $1.25 \times 0.75 \ \mu m$ (Fig. 2). Table 1 summarizes the amount of growth, degree of sporulation, spore mass color, color of the reverse-substrate mycelium, and presence of soluble pigment produced by strain X-14873 on various agar media.

Physiological and Chemical Characteristics

The carbon utilization and other metabolic characteristics of strain X-14873 are shown in Tables 2 and 3. The culture hydrolyzes casein, starch, gelatin, adenine, xanthine, hypoxanthine, tyrosine,

[†] Present address: Smith Kline & French Laboratories, P.O. Box 7929, Philadelphia, PA 19101, U.S.A.

Fig. 1. The structure of antibiotic X-14873A, X-14873H, and X-14873G.









X-14873G

Fig. 2. Streptomyces sp. X-14873.

Smooth spores and a chain of undifferentiated spores on ISP 2 after 14 days of incubation at 28°C. Treatment included glutaraldehyde fixation and critical point drying. Bar represents 1 μ m in length.



and urea by the method of GORDON⁹⁾. As shown in Table 2, strain X-14873 does not grow on cellulose or sucrose. The analysis of the isomers

of diaminopimelic acid in the cell wall of strain X-14873 was carried out according to the method of BECKER *et al.*¹⁰⁾. The cell wall contains the LL-isomer of diaminopimelic acid which, together with the above characteristics, places this organism in the genus *Streptomyces*¹¹⁾. According to BERGEY'S Manual¹²⁾, *Streptomyces* strain X-14873 resembles *S. chrysomallus*, *S. parvus*, and *S. globisporus*. All but *S. globisporus* produce actinomycin. These known cultures possess a yellow-colored spore mass, while strain X-14873 is mainly gray with only a hint of yellow. Strain X-14873 produces a yellow soluble pigment as do the other strains. The carbon utilization patterns are very similar for all the strains. Since the cultures all hydrolyze starch, gelatin, casein, and urea, and decompose adenine, xanthine, hypoxanthine, and tyrosine, it is difficult to distinguish between these cultures based on the above criteria.

Fermentation

Spores of strain X-14873 were used to inoculate a 500-ml Erlenmeyer flask containing 100 ml of inoculum medium having the following composition (in g/liter): tomato pomace 5.0, distiller's soluble 5.0, meat peptone (Type-SB, Marco Development Corp., Hackensack, NJ, U.S.A.) 5.0, debittered dried yeast 5.0, Eclipse N starch (A.E. Staley, Decatur, IL, U.S.A.) 20.0, CaCO₃ 1.0, and K_2HPO_4 1.0. The pH of the medium was adjusted to 7.0 before autoclaving. The inoculated flask was incubated for 96 hours at 28°C on a rotary shaker (5-cm gyration) operating at 250 rpm. One ml of the resulting vegetative growth was used as inoculum to start fermentation in a 500-ml Erlenmeyer flask containing 50 ml of production medium (in g/liter): glycerol 40.0, Cerelose (technical grade glucose)

THE JOURNAL OF ANTIBIOTICS

Agar medium	Amount of growth, degree of sporulation	Spore mass color ^a	Color of reverse- substrate mycelium ^a
Yeast - malt extract (ISP 2) ^b	Moderate to abundant growth; moderate sporulation; brown soluble pigment	c (light gray) where sporulated; 3ni (clove brown) where not sporulated	3ie (camel) and 2gc (light tan) edge
Oatmeal (ISP 3) ^b	Moderate growth; sparse sporulation; yellow soluble pigment	b (oyster white) where sporulated; 2gc (bamboo) where not sporulated	2ec (biscuit)
Inorganic salts - starch (ISP 4) ^b	Abundant growth; well sporulated; hydrolyzes starch; yellow soluble pigment	3cb (sand)	3lc (amber) and 3le (cinnamon)
Glycerol - asparagine (ISP 5) ^b	Moderate growth; moderate sporulation; yellow-brown soluble pigment; hygroscopic	b (oyster white)	3le (cinnamon)
Czapek - Dox (BBL)	Moderate growth; moderate sporulation; yellow soluble pigment; slightly hygroscopic	b (oyster white)	2ic (honey gold)

Table 1. Cultural characteristics of strain X-14873.

^a The color scheme used was Color Harmony Manual, 4th Ed., 1958, Container Corporation of America, Chicago, IL, U.S.A.

^b Media recommended by the International Streptomyces Project¹⁸⁾.

Carbon		Result ^a		
source	X-14873	S. chrysomallus NRRL 225D	<i>S. parvus</i> NRRL B-1455	S. globisporus ATCC 15864
No carbon control	_	_		_
D-Glucose	++	++ YS	++ YS	++ YS
D-Xylose	\pm	+	++ YS	+
L-Arabinose	-[-	+	+ (+)	+
L-Rhamnose	++ YS	++ YS	++	++
D-Fructose	++	++	++	++
D-Galactose	++	++ YS	++ YS	++
Raffinose	\pm	_	_	-
D-Mannitol	++ YS	++ YS	++ YS	++
<i>i</i> -Inositol	±	-	-	—
Salicin	+	\pm	\pm	±
Sucrose	-	—		-
Cellulose	-	-		
Maltose	++	++ YS	++ intense YS	++
Glycerol	++	++	++ intense YS	++
Starch	++	++ YS		++
Ribose	++	++ YS	++ YS	++ YS

Table 2. Comparison of carbon utilization by strain X-14873 and related strains on ISP 9 medium.

^a -: Negative response; $\pm:$ doubtful response; +: more growth than on carbon control but less than on glucose; +(+): positive response, nearly equal to growth on glucose. ++: positive response equal to growth on glucose. YS: yellow soluble pigment.

All tests were performed by the method of SHIRLING and GOTTLIEB¹⁶⁾.

Test	Result ^a	Test	Result ^a
ISP 6, darkening	-	Streptomycin sensitivity	+13 mm
Melanin, ISP 7	_	(10 µg disc)	
ISP 1, darkening	_	Lysozyme sensitivity ^e	+
Gelatin hydrolysis ^b	+	Nitrate reduction ^e	+
Casein hydrolysis ^e	+	Hygroscopic property	+
Starch hydrolysis ^d	+	Antibiotic production	Polyether antibiotics
NaCl (%) tolerance ^c	5		X-14873 A, G, & H;
Growth range temp (°C)	10~26		Actinomycins
Reverse-side pigment	-		X-14873 B, C, & D
Soluble pigment	Yellow		

Table 3. Metabolic characteristics of X-14873.

^a All tests were done at 28°C.

^b Test was performed by the method of SKERMAN¹⁷⁾.

^c Test was performed by the method recommended in GORDON and SMITH¹⁸).

^d Determination was made after growth on agar of Actinomyces broth (Difco) with 0.25% soluble starch by flooding the plates with iodine-KI solution.

^e Test was performed by the method of GORDON¹⁹.

Fig. 3. Time course of X-14873A fermentation in shake flask.

The fermentation was carried out in 500-ml Erlenmeyer flasks containing 50 ml of production medium under the condition described in the text.



2.0, meat peptone (Type-SB, Marco Development Corp., Hackensack, NJ, U.S.A.) 5.0, potato starch 5.0, toasted Nutrisoy (Archer Daniels Midland Co., Decatur, IL, U.S.A.) 5.0, NaCl 5.0, and $CaCO_3$ 2.0. The medium pH was adjusted to 6.4 before sterilization. The fermentation was carried out at 28°C on a rotary shaker (5-cm gyration) operating at 250 rpm.

The amount of antibiotic X-14873A produced in the fermentation broth was determined by ethyl acetate extraction and high pressure liquid chromatography. The analysis was carried out on a reverse phase C_{18} -Nova Pak (15 cm × 4.5 mm, Waters Assoc., Milford, MA, U.S.A.) using a solvent sys-

i z al la	Organism	Minimal i	nhibitory conce (µg/ml) ^a	entration
		X-14873A	X-14873G	X-14873H
G(-) rods	Pseudomonas aeruginosa ATCC 8705	>1,000	>1,000	>1,000
	Proteus vulgaris ATCC 6380	>1,000	>1,000	>1,000
	Escherichia coli ATCC 27856	>1,000	>1,000	>1,000
	Klebsiella pneumoniae ATCC 27858	>1,000	>1,000	>1,000
	Serratia marcescens ATCC 27857	>1,000	>1,000	>1,000
	Acinetobacter calcoaceticus ATCC 10153	>1,000	>1,000	>1,000
G(+) cocci	Streptococcus faecium ATCC 8043	0.23	62.5	3.9
	Staphylococcus aureus ATCC 6538P	3.9	1,000	7.9
	Micrococcus luteus ATCC 9341	7.9	1,000	3.9
G(+) rods	Bacillus megaterium ATCC 8011	7.9	1,000	3.9
	Bacillus sp. E ATCC 27359	0.23	7.9	3.9
	B. subtilis NRRL 558	1.9	>1,000	7.9
	Bacillus sp. TA ATCC 27860	1.9	>1,000	7.9
	Mycobacterium phlei ATCC 355	31.3	>1,000	>1,000
G(+) filament	Streptomyces cellulosae ATCC 3313	15.7	>1,000	62.5
Molds	Paecilomyces varioti ATCC 25820	62.5	>1,000	>1,000
	Penicillium digitatum ATCC 26821	>1,000	>1,000	>1,000
Yeasts	Candida albicans NRRL 477	31.3	>1,000	>1,000
	Saccharomyces cerevisiae ATCC 4226	>1,000	>1,000	>1,000

Table 4. In vitro antimicrobial activity of antibiotic X-14873A, X-14873G, and X-14873H.

^a Lowest two-fold dilution still showing zone of inhibition by an agar well diffusion assay. G(+): Gram-positive; G(-): Gram-negative.

tem composed of acetonitrile and 1% aq acetic acid (85: 15) with a flow rate of 0.9 ml/minute. Antibiotic X-14873A was observed for the flow time at 6 minutes (K' 4.5) using a RI detector (ERC-7510, Erma Optical Works, Tokyo, Japan). Glycerol content in fermentation broth was determined by a colorimetric method¹³. The cell growth was estimated by measuring the packed-mycelia volume in broth by centrifugation at 1,600 rpm for 10 minutes in a graduated glass centrifuge tube (13×130 mm) with a Sorvall GLC-2B bench-top centrifuge. A typical time course of X-14873 is shown in Fig. 3.

Biological and Ionophore Properties

As an antibiotic, X-14873A is mainly active against Gram-positive bacteria and exhibits no activity against all the Gram-negative rods tested. The antimicrobial spectra of X-14873A, X-14873G, and X-14873H are shown in Table 4. It is interesting to note that X-14873H, the descarboxyl molecule of X-14873A, is also active against most of the Gram-positive bacteria tested while the other descarboxyl derivative, X-14873G, is practically inactive. These results indicate that the carboxylic function of the polyether antibiotic molecule is not required for the antimicrobial activity, even though the ionized carboxylic group plays an important role in complexation of cations by carboxylic acid polyether antibiotics¹⁾.

Antibiotic X-14873A also induces a change in the proportion of volatile fatty acids (acetate, propionate, and butyrate) produced in the rumen by increasing the molar proportion of propionate in the rumen fluid. This activity has been implicated as the mechanism for increased feed utilization in ruminants by the polyether antibiotics, lasalocid and monensin¹⁴⁾, and suggests that antibiotic X-14873A is a potential candidate as a ruminant performance enhancer.

X-14873A is structurally similar to lysocellin¹⁵⁾ and exhibits similar ionophorous properties²⁾.

●: Ca²⁺; ▲: Rb⁺.

in Table 5.

Cs+

Fig. 4. Time course of Ca^{2+} and Rb^+ transport by antibiotic X-14873A in a *U*-tube.

The U-tube system described by ASHTON and STEINRAUF²¹⁾ was employed for the assay. Five ml of a CHCl₃ solution of antibiotic $(1 \times 10^{-4} \text{ M})$ were added to a glass U-tube. Two ml of an aqueous Tris-HCl buffer (20 mm, pH 9.5) containing 1 mM [45Ca]calcium chloride or [86Rb]rubidium chloride was placed in one arm of the U-tube and an equal volume of the same buffer solution with unlabelled calcium or rubidium chloride in the other arm. The chloroform phase separating the two aqueous phases was then gently stirred with a magnetic stirrer. The rate of appearance of radioactive calcium or rubidium in the label-free side was determined by counting samples (50 μ l) taken from both aqueous phases with 10 ml Aquasol (New England Nuclear, Boston, Mass., U.S.A.) in an Intertechnique liquid scintillation spectrometer.

0.7 0.6 Transported Ca^{2+} or Rb^+ (μM) 0.5 0.4 0.3 0.2 0.1 0 0 2 4 6 8 10 12 Time (hours)

Displacing cation	Cation radius (Å)	⁸⁶ Rb ⁺ or ⁴⁵ Ca ²⁺ remaining in complex (%)		
		⁸⁶ Rb ⁺	${}^{45}Ca^{2+}$	
None		62.3	77.7	
Mg^{2+}	0.82	47.7	43.7	
Ca ²⁺	1.18	48.0	43.8	
Sr^{2+}	1.12	54.5	39.8	
Ba^{2+}	1.34	39.1	13.7	
Li ⁺	0.68	59.2	72.8	
Na ⁺	0.97	46.4	50.5	
K ⁺	1.33	32.9	33.0	
Rh ⁺	1 47	34 5	35 3	

1.67

Table 5. Displacement of ⁸⁶Rb⁺ or ⁴⁵Ca²⁺ from

X-14873A-cation complex by other cations.

The cation selectivity sequence of X-14873A was determined by the method of LIU and HERMANN²⁰⁾. Two μ mol of antibiotic (in 2 ml of 70% toluene-30% butanol) were equilibrated by shaking for 2 hours at 20°C with 2 μ mol of ⁸⁶RbCl or 1 μ mol of ⁴⁸CaCl₂ (in 1 ml of dimethylglycine-tetramethyl-ammonium hydroxide buffer, 25 mM pH 10.0). Complex formation was measured by liquid scintillation counting of both phases (see Fig. 4).

40.5

43.8

Displacement of labeled cation from the ionophore complexes was determined by the addition of unlabeled cations (2 μ mol of monovalent or 1 μ mol of divalent metal chloride).

It complexes and transports both monovalent and divalent cations as demonstrated in the experiments shown in Table 5 and Fig. 4. The cation binding sequence of X-14873 as determined by an ion-competition experiment (Table 5) is:

 $Ba^{2+} \gg K^+, Rb^+ > Ca^{2+}, Mg^{2+},$

As expected, the descarboxylic derivatives

of X-14873A, *i.e.*, X-14873G and X-14873H, do not bind and transport either rubidium or calcium ions in the two-phase distribution system described

References

- LIU, C.-M.: Microbial aspects of polyether antibiotics: Activity, production, and biosynthesis. In Polyether Antibiotics: Naturally Occurring Acid Ionophores. Vol. 1, Biology. Ed., J. W. WESTLEY, pp. 44~102, Marcel Dekker, Inc., New York, 1982
- 2) LIU, C.-M.; T. E. HERMANN, B. LA T. PROSSER, N. J. PALLERONI, J. W. WESTLEY & P. A. MILLER: X-14766A, a halogen containing polyether antibiotic produced by *Streptomyces malachitofuscus* subsp. *downeyi* ATCC 31547. Discovery, fermentation, biological properties and taxonomy of the producing culture. J. Antibiotics 34: 133~138, 1981
- 3) LIU, C.-M.; T. E. HERMANN, B. LA T. PROSSER, N. J. PALLERONI, J. W. WESTLEY & P. A. MILLER: Novel

polyether antibiotics X-14667A and X-14667B from *Streptomyces cinnamonensis* subsp. *urethanofaciens*. Discovery, fermentation, biological, as well as ionophore properties and taxonomy of the producing culture. J. Antibiotics 34: 1241~1247, 1981

- 4) LIU, C.-M.; T. E. HERMANN, A. DOWNEY, B. LA T. PROSSER, E. SCHILDKNECHT, N. J. PALLERONI, J. W. WESTLEY & P. A. MILLER: Novel polyether antibiotics X-14868A, B, C, and D produced by a Nocardia. Discovery, fermentation, biological as well as ionophore properties and taxonomy of the producing culture. J. Antibiotics 36: 343~350, 1983
- 5) WESTLEY, J. W. (*Ed.*): Polyether Antibiotics: Naturally Occurring Acid Ionophores. Vol. 1, Biology. Marcel Dekker, Inc., New York, 1982
- ÖTAKE, N.; H. NAKAYAMA, H. MIYAMAE, S. SATO & Y. SAITO: X-ray crystal structure of the thallium salt of carriomycin a new polyether antibiotic. Chem. Commun. 1977: 590~591, 1977
- 7) TONE, J.; R. SHIBAKAWA, H. MAEDA, K. INOUE, S. NISHIYAMA, M. ISHIGURO, W. P. CULLEN, J. B. ROUTIEN, L. R. CHAPPEL, C. E. MOPPETT, M. T. JEFFERSON & W. D. CELMER: CP-44,161, a new polycyclic ether antibiotic produced by a new species of *Dactylosporangium*. Program and Abstracts of the 18th Intersci. Conf. on Antimicrob. Agents Chemother., No. 171, Atlanta, Oct. 1~4, 1978
- WESTLEY, J. W.; C.-M. LIU, J. F. BLOUNT, L. TODARO, L. H. SELLO & N. TROUPE: Isolation and characterization of three novel polyether antibiotics and three novel actinomycins as cometabolites of the same *Streptomyces* sp. X-14873, ATCC 31679. J. Antibiotics 39: 1704~1711, 1986
- 9) GORDON, R. E.: The taxonomy of soil bacteria. In Ecology of Soil Bacteria. Eds., T. R. G. GRAY & D. PARKINSON, pp. 293~321, Liverpool University Press, Liverpool, 1967
- BECKER, B.; M. P. LECHEVALIER, R. E. GORDON & H. A. LECHEVALIER: Rapid differentiation between Nocardia and Streptomyces by paper chromatography of whole-cell hydrolysates. Appl. Microbiol. 12: 421~423, 1964
- LECHEVALIER, H. A.; M. P. LECHEVALIER & N. N. GERBER: Chemical composition as a criterion in the classification of actinomycetes. Adv. Appl. Microbiol. 14: 47~72, 1971
- BUCHANAN, R. E. & N. E. GIBBONS (Ed.): BERGEY'S Manual of Determinative Bacteriology. 8th Ed., pp. 727~729, Williams & Wilkins Co., Baltimore, 1974
- LAMBERT, M. & A. C. NEISH: Rapid method for estimation of glycerol in fermentation solution. Can. J. Res. 28B: 83~89, 1950
- 14) RUFF, M. D.: Veterinary application. In Polyether Antibiotics: Naturally Occurring Acid Ionophores. Vol. 1, Biology. Ed., J. W. WESTLEY, pp. 303~332, Marcel Dekker, Inc., New York, 1982
- 15) ÖTAKE, N.; M. KOENUMA, H. KINASHI, S. SATO & Y. SAITO: The crystal and molecular structure of the silver salt of lysocellin, a new polyether antibiotic. Chem. Commun. 1975: 92~93, 1975
- 16) SHIRLING, E. B. & D. GOTTLIEB: Methods for characterization of *Streptomyces* species. Int. J. Syst. Bacteriol. 16: 313~340, 1966
- 17) SKERMAN, V. B. D. (*Ed.*): A Guide to Identification of the Genera of Bacteria. Williams & Wilkins Co., Baltimore, 1967
- 18) GORDON, R. E. & M. M. SMITH: Rapidly growing acid fast bacteria. J. Bacteriol. 66: 41~48, 1953
- GORDON, R. E.: Some criteria for the recognition of Nocardia madurae (Vincent) Blanchard. J. Gen. Microbiol. 45: 355~364, 1966
- 20) LIU, C.-M. & T. E. HERMANN: Characterization of ionomycin as a calcium ionophore. J. Biol. Chem. 253: 5892~5894, 1978
- ASHTON, R. & L.K. STEINRAUF: Thermodynamic consideration of the ion transport antibiotics. J. Mol. Biol. 49: 547~556, 1970